

Absorption correction:
numerical (XRED; Stoe
& Cie, 1997a)
 $T_{\min} = 0.031$, $T_{\max} = 0.233$
9080 measured reflections
4196 independent reflections

Refinement

Refinement on F^2
 $R[F^2 > 2\sigma(F^2)] = 0.032$
 $wR(F^2) = 0.044$
 $S = 1.844$
4196 reflections
144 parameters
 $w = 1/[\sigma^2(F_o^2)]$
 $(\Delta/\sigma)_{\max} = 0.001$
 $\Delta\rho_{\max} = 1.273 \text{ e } \text{\AA}^{-3}$
 $\Delta\rho_{\min} = -1.865 \text{ e } \text{\AA}^{-3}$

$R_{\text{int}} = 0.048$
 $\theta_{\max} = 28.06^\circ$
 $h = -11 \rightarrow 10$
 $k = -11 \rightarrow 11$
 $l = -18 \rightarrow 18$

Extinction correction:
SHELXL93 (Sheldrick,
1993)
Extinction coefficient:
0.00139 (11)
Scattering factors from
International Tables for
Crystallography (Vol. C)

Table 1. Selected geometric parameters (\AA , $^\circ$)

Br1—C1	1.894 (5)	C1—C6	1.374 (7)
Br2—C2	1.871 (5)	C1—C2	1.407 (7)
Br3—C3	1.908 (5)	C2—C3	1.408 (7)
Br4—C4	1.902 (5)	C3—C4	1.380 (7)
Br5—C5	1.885 (6)	C4—C5	1.404 (7)
Br6—C8	1.899 (6)	C5—C6	1.394 (6)
Br7—C9	1.891 (5)	C7—C12	1.374 (8)
Br8—C10	1.895 (5)	C7—C8	1.405 (7)
Br9—C11	1.886 (6)	C8—C9	1.390 (7)
Br10—C12	1.898 (5)	C9—C10	1.404 (8)
O—C7	1.389 (6)	C10—C11	1.390 (7)
O—C6	1.406 (5)	C11—C12	1.408 (8)
C1—C6—C5	122.0 (4)	C12—C7—O	125.8 (5)
C1—C6—O	116.1 (4)	C12—C7—C8	119.8 (5)
C5—C6—O	121.9 (5)	O—C7—C8	114.1 (5)
C7—O—C6—C1	125.2 (5)	C6—O—C7—C12	-39.0 (6)
C7—O—C6—C5	-56.0 (7)	C6—O—C7—C8	146.4 (4)

All Br atoms were refined with anisotropic displacement parameters. The large positive and negative differences are located near the Br atoms. Refining the C atoms with anisotropic displacement parameters yielded some of them as slightly non-positive definite, but with no significant improvement in the residual factors. Since the thermal vibrations were rather small due to the low-temperature experiment, it was decided to use isotropic C atoms in the structure model in order to avoid non-physical displacement parameters.

Data collection: EXPOSE (Stoe & Cie, 1997b). Cell refinement: CELL (Stoe & Cie, 1997c). Data reduction: INTEGRATE (Stoe & Cie, 1997d). Program(s) used to solve structure: SHELXS86 (Sheldrick, 1990). Program(s) used to refine structure: SHELXL93 (Sheldrick, 1993). Molecular graphics: DIAMOND (Bergerhoff, 1996).

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Supplementary data for this paper are available from the IUCr electronic archives (Reference: KA1314). Services for accessing these data are described at the back of the journal.

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L-Alanyl-L-phenylalanine-2-propanol (1/2) (α -form), L-valyl-L-phenylalanine-2-propanol (1/1) and L-leucyl-L-phenylalanine-2-propanol (1/1) (β -form)

CARL HENRIK GÖRBITZ

Department of Chemistry, University of Oslo, PO Box 1033 Blindern, N-0315 Oslo, Norway. E-mail: c.h.gorbitz@kjemi.uio.no

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Abstract

Small crystals of L-Ala-L-Phe-2-propanol (1/2), $\text{C}_{12}\text{H}_{16}\text{N}_2\text{O}_3 \cdot 2\text{C}_3\text{H}_8\text{O}$, L-Val-L-Phe-2-propanol (1/1), $\text{C}_{14}\text{H}_{20}\text{N}_2\text{O}_3 \cdot \text{C}_3\text{H}_8\text{O}$, and L-Leu-L-Phe-2-propanol (1/1), $\text{C}_{15}\text{H}_{22}\text{N}_2\text{O}_3 \cdot \text{C}_3\text{H}_8\text{O}$, were obtained after considerable effort. The three structures have intricate packing interactions with up to four peptide molecules in the asymmetric unit and variable hydrogen-bond connectivities for the carboxylate groups. For each peptide, two or more different crystal forms were obtained in the crystallization experiments.

Comment

For dipeptides with two hydrophobic residues our previous (Görbitz, 1997, and references therein) and ongoing work has identified two different modes of

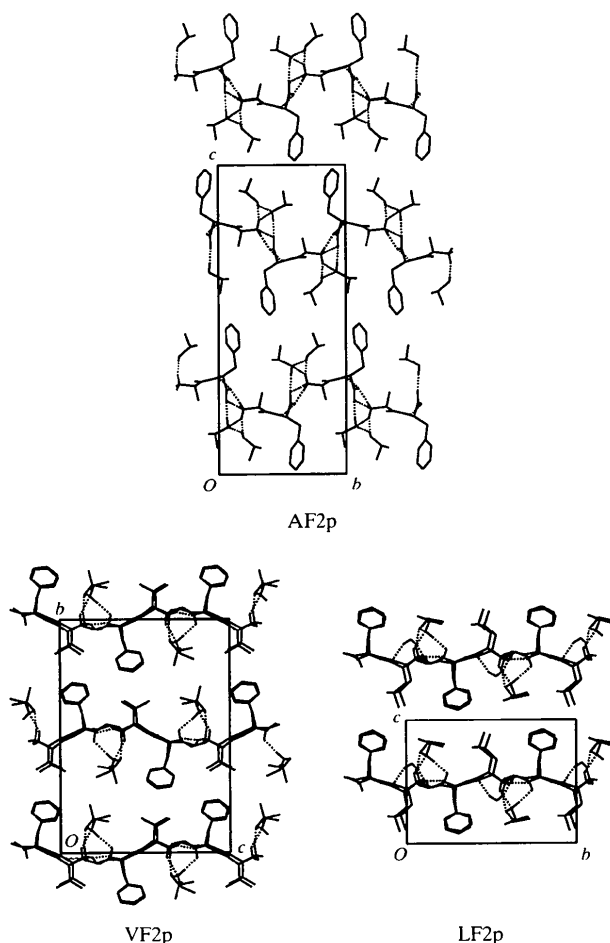


Fig. 2. The unit cell and crystal packing for the three title compounds. H atoms bonded to C atoms have been omitted for clarity. Horizontal hydrophilic layers and hydrophobic layers are seen edge-on.

layer, and neighbouring hydrophilic layers are related by the crystallographic screw axis. Additionally, a non-crystallographic screw axis parallel to the c axis at $x = \frac{3}{8}$ and $y = 0$ very accurately relates A to C and B to D (r.m.s. deviations 0.039 and 0.062 Å, respectively) and solvent molecules E to G and F to H . A comprehensive discussion of the symmetry and systematic absences (see below) for this special monoclinic system is given by Görbitz & Torgersen (1999) for the isomorphous and completely analogous structure of LVe.

Pseudotranslational symmetry along the a axis is also observed for LF2p, with a heavy-atom r.m.s. deviation of 0.218 Å for the best fit between the two molecular geometries. The peptide conformations are very similar to those observed in VF2p, as is evident from Fig. 1, with large values for ψ_T (150–159°) and a *gauche*⁺ orientation for the L-Phe side chain. As for VF2p, neighbouring solvent molecules along the a axis have somewhat different orientations (Fig. 1c). Furthermore, it can be seen from Fig. 1 that the solvent mol-

ecules have roughly equivalent positions (albeit different orientations) in both structures. The exception is the partially occupied [0.102(5)] solvent water molecule, which is present in LF2p only. It sits in a small cavity that is created when the L-Phe side chain is reoriented to compensate for the lack of side-chain branching at C^β for residue 1 as Val (VF2p) is replaced by Leu (LF2p).

The hydrogen-bond patterns of AF2p and LF2p (Fig. 3) are typical for hydrophobic dipeptides, in that each amino group donates two H atoms to carboxylate groups, forming head-to-tail chains, while the third amino-H atom is donated to the hydroxyl group of a cocrystallized alcohol molecule. Thus, the solvent molecules are not only included to fill cavities in the hydrophobic layers, but are indispensable parts of the hydrogen-bonding network in these structures. The AF2p pattern is identical to the pattern observed in the structure of LVm (Görbitz & Torgersen, 1999) if the 2-propanol molecule B is removed and the hydroxyl group of the 2-propanol molecule C is rotated to form an interaction with the peptide carbonyl oxygen O1A. Such a pattern is also observed in L-Leu-L-Tyr (LY; Krause *et al.*, 1993), in which the hydroxyl group comes from the Tyr side chain and not from a cocrystallized alcohol molecule. The three structures share the short a axis [AF2p 5.4825(4), LVm 5.2890(1) and LY 5.644(1) Å] while, as for several other layered dipeptides, a second axis (b for all) is close to 12 Å, which follows from the orientation of the peptide main chain. The difference between the 11.4872(1) Å c axis for LVm and the 29.4245(19) Å c axis for AF2p reflects not only a doubling upon transition from space group $P2_1$ to $P2_12_12_1$, but also a further increase due to the large Phe side chain in AF2p. The larger separation between hydrophilic layers in AF2p means that the cavity filled by the methanol molecule in LVm becomes larger, and is instead filled by the 2-propanol molecule C . The second solvent molecule in AF2p (B) fills the void that

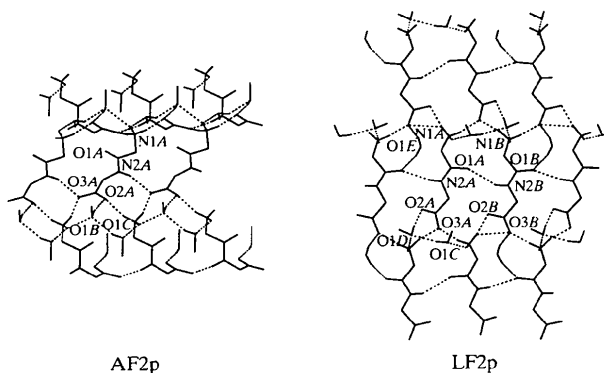
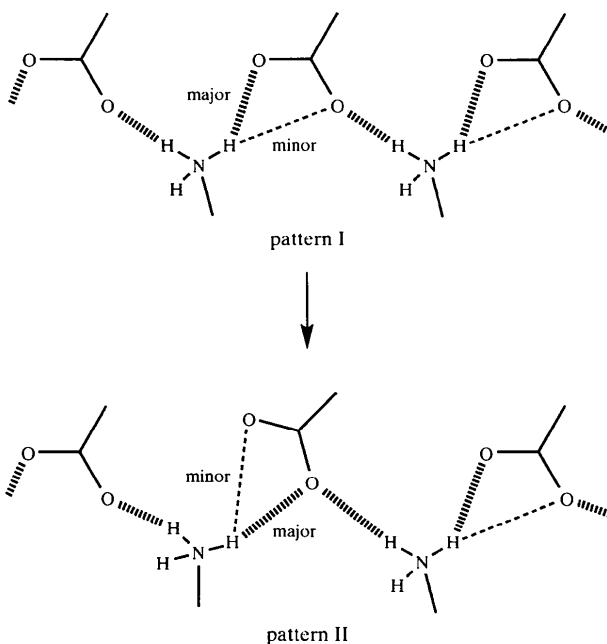


Fig. 3. The hydrogen-bonding pattern (dashed lines) in the crystal structures of AF2p and LF2p. All peptide side chains and H^α atoms have been removed for clarity; for the alcohol molecules, only C—O—H remains. Essential atom labels have been indicated. Hydrogen bonds involving LF2p water molecules with low occupancy are shown as dotted lines.

is generated when Leu in the LVm structure is replaced by Ala.

The hydrogen-bond patterns of VF2p and LF2p (Fig. 3) are almost identical, except for interactions involving the partially occupied LF2p solvent water molecule. The lengths of the *a* axes are 9.8958 (2) Å for VF2p and 10.0858 (2) Å for LF2p, which are close to the lengths of the corresponding axes for LVe and LLmp [11.0112 (1) and 10.3179 (1) Å, respectively; Görbitz, 1999]. The doubling of the *a*-axis lengths compared with AF2p and LVm results from a shift in the basic hydrogen-bond connectivity for every second carboxylate group (see Scheme below).



The major components in both types of asymmetric carboxylate interactions have H···O distances in the range 1.82–1.88 Å, while H···O distances for the long minor components are in the range 2.56–2.64 Å.

An important difference between the two hydrogen-bond patterns shown in Fig. 3 is that AF2p (and thus also LVm, LVe and LV2p) has a characteristic >N—H···O interaction between the peptide group and the terminal-C carboxylate group. The carbonyl O atom, on the other hand, is not an H-atom acceptor and the C=O bond distance is short [1.201 (5) Å]. In contrast, the amide-H atoms in VF2p and LF2p are involved in alternating weak/very weak hydrogen bonds to peptide carbonyl groups, with additional weak/very weak C^α—H···O=C interactions varying in the opposite sequence (Tables 4 and 6). An inherently high stability for carboxylate pattern II is suggested by the fact that it can be integrated into both of these hydrogen-bond patterns, as observed in the structures of LVe and LV2p, and VF2p and LF2p, respectively.

As described in the *Experimental* section below, two or more different types of crystals were obtained in the crystallization experiments for all three dipeptides studied. The structure of the LF2p β -form (plate-shaped crystals) presented here is, judged by the cell dimensions, very closely related to the structure of the α -form [needle-shaped crystals: *a* = 9.9552 (3), *b* = 16.3894 (5), *c* = 23.7291 (2) Å, and β = 90.0396 (16)°]. The situation is more complicated for the relationship between the two crystal forms of AF2p for which data have been recorded. The cell volume for the AF2p β -form [*a* = 12.3485 (1), *b* = 13.5161 (1) and *c* = 28.9248 (4) Å] is 4828 Å³, or 2.42 times the cell volume of the α -form. This means that the asymmetric unit of β -AF2p either contains more than two solvent molecules or possibly two independent peptide molecules. The latter alternative gives a calculated density of 1.301 Mg m⁻³, which is high for hydrophobic dipeptides. This would leave no room for additional 2-propanol molecules, and probably not even solvent water molecules. Accordingly, such a structure would deviate from other known structures of hydrophobic dipeptides. We attempted to measure the crystal density experimentally by flotation, but no reliable results were obtained since only a limited number of very small crystal specimens was available. Attempts to solve this structure will continue.

Experimental

The title compounds were purchased from Sigma and used as received. About 0.1–0.3 mg of each peptide was dissolved in 30 μ l water with subsequent vapour diffusion of 2-propanol (or other alcohol) into the aqueous solution at room temperature. The crystallizations for L-Ala-L-Phe uniformly produced very thin needles unsuitable for diffraction purposes. One test tube from these experiments was accidentally left in the laboratory and was, upon checking four months later, found to contain small blocks. Several crystals were tested and found to belong to two different forms, *i.e.* an orthorhombic α form and a β form of unknown crystal system with roughly twice the unit-cell volume. The crystallization experiment was repeated, but this time only a few blocks were obtained, all of the β -modification. Additionally, some rod-like needles were formed which appear to be distinct from the other crystals. We were not able to collect data on any of these rods. The experiments and the results for L-Val-L-Phe are very similar to those for L-Ala-L-Phe. After four months, the initial very thin needles had been converted to plate-shaped crystals. Alas, the crystals proved to be very unstable and decayed instantly when exposed to air. One small single crystal, however, remained stable suggesting it was of a different modification. Data were collected from this specimen. The crystallization was repeated under the same conditions and after a period of months, small crystals had again appeared. This time all the crystals were of the stable modification, but were not larger or better than that obtained earlier. Like L-Ala-L-Phe and L-Val-L-Phe, L-Leu-L-Phe at first yielded only very thin needles (α -form) in the crystallizations. After numerous attempts, a needle just large enough for data collection was obtained.

Further experiments revealed that a second crystal form (β -form) could be obtained with low initial peptide concentration, i.e. 0.08 mg in 30 μ l water (concentrations below 0.06 mg in 30 μ l did not produce and precipitate). In the first series of crystallizations, only 2-propanol was used as precipitating agent. Later, similar experiments were carried out with seven other alcohols. Generally, very thin needles were obtained for all three peptides. The exceptions were ethanol batches of L-Ala-L-Phe and L-Val-L-Phe, which also yielded some extremely thin plates. Attempts to collect data on these crystals were unsuccessful.

Compound AF2p*Crystal data*C₁₂H₁₆N₂O₃·2C₃H₈O $M_r = 356.46$

Orthorhombic

P2₁2₁2₁ $a = 5.4835$ (4) Å $b = 12.3418$ (8) Å $c = 29.4245$ (19) Å $V = 1991.3$ (2) Å³ $Z = 4$ $D_x = 1.189$ Mg m⁻³ D_m not measured*Data collection*

Siemens SMART CCD diffractometer

Sets of exposures each taken over 0.2° ω rotation scans

Absorption correction: empirical (SADABS; Sheldrick, 1996)

 $T_{\min} = 0.981$, $T_{\max} = 0.997$

10 243 measured reflections

3490 independent reflections

*Refinement*Refinement on F^2 $R[F^2 > 2\sigma(F^2)] = 0.076$ $wR(F^2) = 0.203$ $S = 1.096$

3490 reflections

234 parameters

H atoms constrained

 $w = 1/[\sigma^2(F_o^2) + (0.0959P)^2 + 1.2701P]$ where $P = (F_o^2 + 2F_c^2)/3$ Mo $K\alpha$ radiation $\lambda = 0.71073$ Å

Cell parameters from 4425 reflections

 $\theta = 2-25^\circ$ $\mu = 0.086$ mm⁻¹ $T = 150$ (2) K

Block

0.22 × 0.06 × 0.03 mm

Colourless

2524 reflections with $I > 2\sigma(I)$ $R_{\text{int}} = 0.072$ $\theta_{\text{max}} = 25.03^\circ$ $h = -6 \rightarrow 6$ $k = -14 \rightarrow 14$ $l = -35 \rightarrow 30$

Intensity decay: none

 $(\Delta/\sigma)_{\text{max}} = 0.021$ $\Delta\rho_{\text{max}} = 0.34$ e Å⁻³ $\Delta\rho_{\text{min}} = -0.31$ e Å⁻³

Extinction correction: none

Scattering factors from *International Tables for Crystallography* (Vol. C)

Table 1. Selected geometric parameters (Å, °) for AF2p

O1A—C3A	1.201 (5)	N1A—C1A	1.484 (6)
O2A—C12A	1.231 (5)	N2A—C3A	1.363 (5)
O3A—C12A	1.275 (5)		
N1A—C1A—C3A—N2A	159.8 (4)		
C1A—C3A—N2A—C4A	171.4 (4)		
C3A—N2A—C4A—C12A	-77.6 (5)		
N2A—C4A—C12A—O2A	-19.5 (5)		
N2A—C4A—C12A—O3A	162.6 (4)		
N2A—C4A—C5A—C6A	-72.2 (5)		
C4A—C5A—C6A—C7A	127.6 (5)		

Table 2. Hydrogen-bonding geometry (Å, °) for AF2p

D—H...A	D—H	H...A	D...A	D—H...A
N1A—H1A...O1C'	0.91	2.26	2.874 (5)	124
N1A—H1A...O1B''	0.91	2.37	3.030 (5)	130
N1A—H2A...O2A'	0.91	1.98	2.864 (5)	162
N1A—H3A...O3A''	0.91	1.88	2.761 (5)	161
N2A—H4A...O3A'''	0.88	2.17	2.945 (4)	146
O1B—H1B...O2A	0.85	1.94	2.765 (4)	166
O1C—H1C...O1B	0.85	2.12	2.857 (5)	147

Symmetry codes: (i) $1-x, \frac{1}{2}+y, \frac{3}{2}-z$; (ii) $-x, \frac{1}{2}+y, \frac{3}{2}-z$; (iii) $1+x, y, z$.**Compound VF2p***Crystal data*C₁₄H₂₀N₂O₃·C₃H₈O $M_r = 324.41$

Monoclinic

P2₁ $a = 9.8958$ (2) Å $b = 22.3517$ (6) Å $c = 16.4571$ (3) Å $\beta = 90.023$ (1)° $V = 3640.11$ (14) Å³ $Z = 8$ $D_x = 1.184$ Mg m⁻³ D_m not measured*Data collection*

Siemens SMART CCD diffractometer

Sets of exposures each taken over 0.3° ω rotation scans

Absorption correction: empirical (SADABS; Sheldrick, 1996)

 $T_{\min} = 0.979$, $T_{\max} = 0.997$

28 008 measured reflections

12 717 independent reflections

reflections

*Refinement*Refinement on F^2 $R[F^2 > 2\sigma(F^2)] = 0.080$ $wR(F^2) = 0.139$ $S = 1.018$

12 717 reflections

683 parameters

H atoms constrained

 $w = 1/[\sigma^2(F_o^2) + (0.0235P)^2]$ where $P = (F_o^2 + 2F_c^2)/3$ $(\Delta/\sigma)_{\text{max}} = 0.003$ Mo $K\alpha$ radiation $\lambda = 0.71069$ Å

Cell parameters from 7050 reflections

 $\theta = 2-25^\circ$ $\mu = 0.084$ mm⁻¹ $T = 150$ (2) K

Plate

0.25 × 0.10 × 0.04 mm

Colourless

7172 reflections with

 $I > 2\sigma(I)$ $R_{\text{int}} = 0.102$ $\theta_{\text{max}} = 25.03^\circ$ $h = -11 \rightarrow 11$ $k = -26 \rightarrow 26$ $l = -19 \rightarrow 19$

Intensity decay: none

 $\Delta\rho_{\text{max}} = 0.29$ e Å⁻³ $\Delta\rho_{\text{min}} = -0.30$ e Å⁻³

Extinction correction:

SHELXL97 (Sheldrick, 1997a)

Extinction coefficient:

0.0033 (3)

Scattering factors from

International Tables for Crystallography (Vol. C)

Table 3. Selected geometric parameters (Å, °) for VF2p

O1A—C5A	1.255 (4)	O1B—C5B	1.236 (4)
O2A—C14A	1.262 (5)	O2B—C14B	1.245 (5)
O3A—C14A	1.260 (4)	O3B—C14B	1.265 (5)
N1A—C1A	1.507 (4)	N1B—C1B	1.510 (4)
N2A—C5A	1.327 (5)	N2B—C5B	1.350 (5)
N1A—C1A—C5A—N2A	132.3 (5)		
C1A—C5A—N2A—C6A	172.9 (4)		
C5A—N2A—C6A—C14A	-150.8 (5)		
N2A—C6A—C14A—O2A	20.0 (7)		
C7A—C6A—C14A—O3A	-35.3 (7)		
N1A—C1A—C2A—C3A	178.0 (4)		

N1A—C1A—C2A—C4A	−58.3 (5)
N2A—C6A—C7A—C8A	61.1 (5)
C6A—C7A—C8A—C9A	98.2 (7)
N1B—C1B—C5B—N2B	121.9 (5)
C1B—C5B—N2B—C6B	172.5 (4)
C5B—N2B—C6B—C14B	−159.9 (6)
N2B—C6B—C14B—O2B	−11.7 (7)
N2B—C6B—C14B—O3B	171.8 (6)
N1B—C1B—C2B—C3B	−175.6 (5)
N1B—C1B—C2B—C4B	−53.0 (6)
N2B—C6B—C7B—C8B	61.1 (5)
C6B—C7B—C8B—C9B	94.2 (7)
N1C—C1C—C5C—N2C	130.9 (5)
C6C—N2C—C5C—C1C	173.9 (4)
C5C—N2C—C6C—C14C	−149.5 (5)
N2C—C6C—C14C—O2C	19.5 (6)
N2C—C6C—C14C—O3C	−165.7 (5)
N1C—C1C—C2C—C3C	179.3 (4)
N1C—C1C—C2C—C4C	−57.0 (5)
N2C—C6C—C7C—C8C	62.1 (5)
C6C—C7C—C8C—C9C	96.4 (7)
N1D—C1D—C5D—N2D	122.3 (5)
C6D—N2D—C5D—C1D	172.9 (4)
C5D—N2D—C6D—C14D	−159.3 (6)
N2D—C6D—C14D—O2D	−8.9 (7)
N2D—C6D—C14D—O3D	171.5 (6)
N1D—C1D—C2D—C3D	−172.1 (5)
N1D—C1D—C2D—C4D	−49.5 (6)
N2D—C6D—C7D—C8D	61.8 (5)
C6D—C7D—C8D—C9D	91.1 (7)

Data collection

Siemens SMART CCD diffractometer	9829 reflections with $I > 2\sigma(I)$
Sets of exposures each taken over 0.6° ω rotation scans	$R_{\text{int}} = 0.075$ $\theta_{\text{max}} = 37.78^\circ$
Absorption correction: empirical (SADABS; Sheldrick, 1996)	$h = -17 \rightarrow 17$ $k = -27 \rightarrow 26$ $l = -17 \rightarrow 20$
$T_{\text{min}} = 0.964$, $T_{\text{max}} = 0.996$	Intensity decay: none
28 092 measured reflections	
17 153 independent reflections	

Refinement

Refinement on F^2	$(\Delta/\sigma)_{\text{max}} = 0.014$
$R[F^2 > 2\sigma(F^2)] = 0.079$	$\Delta\rho_{\text{max}} = 0.41 \text{ e } \text{Å}^{-3}$
$wR(F^2) = 0.169$	$\Delta\rho_{\text{min}} = -0.27 \text{ e } \text{Å}^{-3}$
$S = 1.050$	Extinction correction: none
17 153 reflections	Scattering factors from <i>International Tables for Crystallography</i> (Vol. C)
447 parameters	
H atoms constrained	
$w = 1/[\sigma^2(F_o^2) + (0.0381P)^2 + 0.5370P]$	
where $P = (F_o^2 + 2F_c^2)/3$	

Table 4. Hydrogen-bonding geometry (Å, °) for VF2p, molecules A, B, E and F

D—H...A	D—H	H...A	D...A	D—H...A
N1A—H1A...O2D ⁱ	0.91	1.84	2.713 (6)	159
N1A—H2A...O3C ⁱⁱ	0.91	1.94	2.786 (7)	154
N1A—H3A...O1F ⁱⁱⁱ	0.91	1.97	2.837 (7)	160
N2A—H4A...O2A	0.88	2.44	2.646 (4)	94
N2A—H4A...O1B	0.88	2.25	3.101 (5)	163
C1A—H5A...O1B	1.00	2.44	3.358 (6)	152
C6A—H6A...O2B ⁱⁱⁱ	1.00	2.40	3.233 (6)	141
N1B—H1B...O3C ⁱ	0.91	1.87	2.765 (6)	167
N1B—H2B...O3D ⁱ	0.91	1.84	2.733 (6)	167
N1B—H3B...O1E	0.91	2.19	2.965 (7)	143
N2B—H4B...O2B	0.88	2.29	2.653 (4)	104
N2B—H4B...O1A ^{iv}	0.88	2.49	3.276 (5)	150
C1B—H5B...O1A ^{iv}	1.00	2.23	3.171 (6)	156
C6B—H6B...O2A	1.00	2.30	3.256 (7)	160
O1E—H1E...O2C ⁱ	0.84	1.84	2.675 (6)	172
O1F—H1F...O1E	0.84	2.07	2.896 (7)	168

Symmetry codes: (i) $x, y, z - 1$; (ii) $1 + x, y, z - 1$; (iii) $1 + x, y, z$; (iv) $x - 1, y, z$.

Compound LF2p

Crystal data

C₁₅H₂₂N₂O₃·C₃H₈O $M_r = 338.44$

Monoclinic

 $P2_1$ $a = 10.0858 (2) \text{ Å}$ $b = 16.3754 (2) \text{ Å}$ $c = 11.7987 (2) \text{ Å}$ $\beta = 91.698 (1)^\circ$ $V = 1947.81 (6) \text{ Å}^3$ $Z = 4$ $D_x = 1.154 \text{ Mg m}^{-3}$ D_m not measuredMo $K\alpha$ radiation $\lambda = 0.71073 \text{ Å}$

Cell parameters from 8192

reflections

 $\theta = 2-35^\circ$ $\mu = 0.081 \text{ mm}^{-1}$ $T = 150 (2) \text{ K}$

Plate

 $0.45 \times 0.35 \times 0.05 \text{ mm}$

Colourless

Table 5. Selected geometric parameters (Å, °) for LF2p

O1A—C6A	1.235 (2)	O1B—C6B	1.236 (2)
O2A—C15A	1.243 (2)	O2B—C15B	1.251 (2)
O3A—C15A	1.274 (2)	O3B—C15B	1.260 (2)
N1A—C1A	1.498 (2)	N1B—C1B	1.495 (3)
N2A—C6A	1.337 (2)	N2B—C6B	1.341 (2)
N1A—C1A—C6A—N2A	127.86 (18)		
C1A—C6A—N2A—C7A	175.34 (17)		
C6A—N2A—C7A—C15A	−152.55 (18)		
N2A—C7A—C15A—O2A	17.5 (2)		
N2A—C7A—C15A—O3A	−164.40 (16)		
N1A—C1A—C2A—C3A	172.01 (17)		
C1A—C2A—C3A—C4A	−178.0 (2)		
C1A—C2A—C3A—C5A	59.0 (3)		
N2A—C7A—C8A—C9A	65.8 (2)		
C7A—C8A—C9A—C10A	98.3 (2)		
N1B—C1B—C6B—N2B	124.93 (18)		
C1B—C6B—N2B—C7B	176.04 (18)		
C6B—N2B—C7B—C15B	−158.97 (18)		
N2B—C7B—C15B—O2B	−5.9 (3)		
N2B—C7B—C15B—O3B	175.00 (19)		
N1B—C1B—C2B—C3B	174.51 (16)		
C1B—C2B—C3B—C4B	177.79 (18)		
C1B—C2B—C3B—C5B	54.6 (3)		
N2B—C7B—C8B—C9B	62.1 (2)		
C7B—C8B—C9B—C10B	96.6 (3)		

Table 6. Hydrogen-bonding geometry (Å, °) for LF2p

D—H...A	D—H	H...A	D...A	D—H...A
N1A—H1A...O2B ⁱ	0.91	1.81	2.715 (2)	170
N1A—H2A...O3A ⁱⁱ	0.91	1.90	2.791 (2)	166
N1A—H3A...O1C ⁱⁱⁱ	0.91	2.15	2.899 (3)	140
N2A—H4A...O2A	0.88	2.42	2.647 (2)	95
N2A—H4A...O1B	0.88	2.29	3.151 (2)	168
C1A—H5A...O1B	1.00	2.58	3.497 (2)	153
C7A—H6A...O2B ⁱⁱⁱ	1.00	2.48	3.323 (2)	141
N1B—H1B...O3A ⁱ	0.91	1.83	2.719 (2)	165
N1B—H2B...O3B ⁱ	0.91	1.82	2.722 (2)	172
N1B—H3B...O1D ⁱ	0.91	2.17	2.854 (2)	131
N2B—H4B...O2B	0.88	2.26	2.619 (2)	105
N2B—H4B...O1A ^{iv}	0.88	2.57	3.372 (2)	152
C1B—H5B...O1A ^{iv}	1.00	2.20	3.180 (2)	165

C7B—H6B···O2A	1.00	2.32	3.288 (2)	164
O1C—H1C···O1D	0.85	2.07	2.895 (2)	169
O1D—H1D···O2A	0.85	1.87	2.700 (2)	172
O1E—H1E···O3B	0.85	2.10	2.911 (12)	159
O1E—H2E···O1B'	0.85	2.13	2.933 (10)	159

Symmetry codes: (i) $1-x, \frac{1}{2}+y, 1-z$; (ii) $-x, \frac{1}{2}+y, 1-z$; (iii) $x-1, y, z$; (iv) $1+x, y, z$; (v) $1-x, y-\frac{1}{2}, 1-z$.

The structures of AF2p (α -form) and LF2p (β -form) were solved directly using *SHELXTL* (Sheldrick, 1997b). The cell dimensions are rather similar to those of LVe ($a = 11.01$, $b = 23.52$, $c = 12.26$ Å and $\beta = 90.01^\circ$; Görbitz & Torgersen, 1999) and the unorthodox systematic absences are the same: $h00$, $h = 4n + 2$ absent; $0k0$, $k = 2n + 1$ absent; $00l$, $l = 2n + 1$ absent. The two structures proved to be isomorphous, and the structure of VF2p was solved in the same indirect manner as described previously for LVe (Görbitz & Torgersen, 1999). As observed for LVe (Görbitz & Torgersen, 1999), the VF2p crystal used for data collection was a TLQS (twin-lattice quasi-symmetry) twin (Giacovazzo *et al.*, 1992), which was effectively treated as TLS (twin-lattice symmetry) since all reflections contained intensity from both twin components. Crystal twinning for a monoclinic system emulating orthorhombic was handled by the *SHELXTL* command *TWIN* 1 0 0 0 -1 0 0 0 -1. The fractions of the two components are 0.576 (2) and 0.424 (2). Pairs of peptide molecules related by non-crystallographic screw axes were connected by tight *SAME* 0.0002 0.0004 commands, constraining equivalent bond lengths and bond angles (but not torsion angles) to be almost similar. This procedure was tested and discussed for refinement of LVe (Görbitz & Torgersen, 1999). Furthermore, U_{ii} ($i = 1, 2$ or 3) and U_{12} values are the same for A and C, and for B and D, while U_{13} and U_{23} values for C and D were constrained to be $-U_{13}$ and $-U_{23}$ for the corresponding atoms in A and B, respectively. Finally, milder *SAME* 0.005 0.008 restraints were used for the geometries of the four 2-propanol molecules. A loose *SAME* 0.01 0.01 restraint was also used for bond lengths and bond angles in the two 2-propanol molecules of AF2p.

For all compounds, data collection: *SMART* (Siemens, 1995); cell refinement: *SAINT* (Siemens, 1995); data reduction: *SAINT*; program(s) used to solve structures: *SHELXTL*; program(s) used to refine structures: *SHELXTL*; molecular graphics: *SHELXTL*; software used to prepare material for publication: *SHELXTL*.

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Supplementary data for this paper are available from the IUCr electronic archives (Reference: OS1078). Services for accessing these data are described at the back of the journal.

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- 4,8-Dibenzyl-2',4',6'-trinitrospiro[1,3-dioxazulonium-2,1'-cyclohexadienide]**
- OLEG YA. BORBULEVYCH,^a MICHAIL YU. ANTIPIN^a AND LEV P. OLEKHOVICH^b
- ^aA. N. Nesmeyanov Institute of Organoelement Compounds of the Russian Academy of Sciences, 28 Vavilov St, Moscow 117813, Russia, and ^bDepartment of Chemistry, Postov State University, 7 Zorge St, Rostov-on-Don 344090, Russia. E-mail: oleg@xrlab.ineos.ac.ru
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Abstract

In molecules of the title compound, C₂₇H₁₉N₃O₈, the nitro groups have almost equivalent geometries. Similar observations were made for some related anionic σ complexes. However, equalization of bond lengths within the conjugated system of the cyclohexadienide ring is stronger than that in other Meisenheimer complexes.

Comment

The synthetic chemistry of electron-deficient aromatics and heteroaromatics is circumscribed by the two major mechanisms of nucleophilic aromatic substitution, *i.e.* SNAr and Vicarious Nucleophilic Substitution (VNS; Artamkina *et al.*, 1982; Bunzel *et al.*, 1995). These displacement reactions form the backbone of numerous important syntheses of pharmaceuticals and potential drugs and several other classes of bioactive agents. The key intermediate in both the SNAr and VNS mechanisms is a negatively charged σ complex commonly termed a Meisenheimer complex.

Dipolar spirocyclic Meisenheimer complexes with tropylium cations are a special class of such complexes and, in general, can be considered as a new class of heterocyclic compounds. In addition, these compounds are of great interest because their formation involves acylotropic rearrangements (Knyazev & Drozd, 1995; Kurbatov *et al.*, 1997).